

# Printing Considerations: Cells, Biologics

NIH Center for Engineering Complex Tissues (CECT)  
June 8, 2018

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Assistant Director, CECT  
University of Maryland




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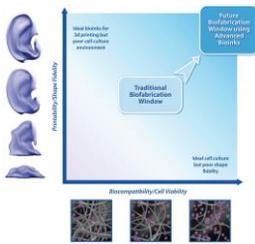
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## Biofabrication window



Kyle et al., Adv. Healthcare Mater., 2017, 6, 1700264




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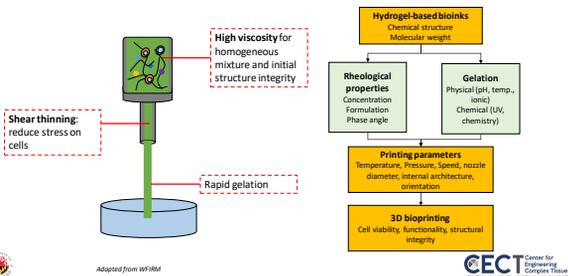
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## Important Bioink properties for extrusion



Adapted from WIBM




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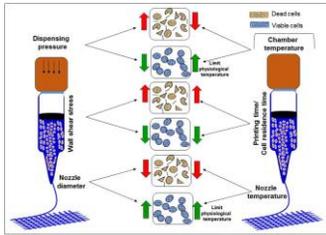
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### Impact of extrusion conditions



Adapted from Biofabrication Workshop, Rice University

Anwar and Tan, Microfluids, 2014




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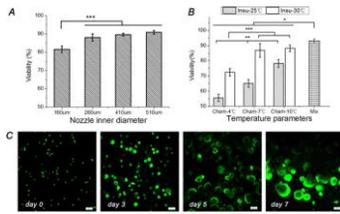
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### Impact of extrusion conditions



Adapted from Biofabrication Workshop, Rice University

Ouyang et al. Biofabrication, 2015




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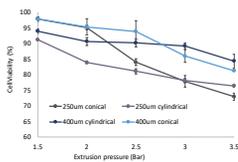
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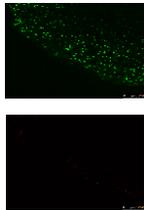
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### Impact of extrusion conditions

- Cell viability in printed fibrin



Slides courtesy Ms. Pinar, TEBL




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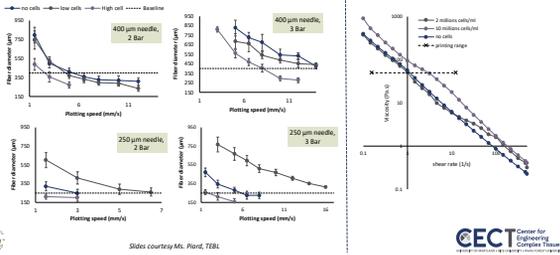
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### Impact of extrusion conditions



### Considerations: Bioink

#### Materials

| ID     | LA/GA Ratio | Molecular Weight | End Cap | Simplified Code |
|--------|-------------|------------------|---------|-----------------|
| PLGA 1 | 50:50       | 9730Da           | ester   | 0.5-10kD-ester  |
| PLGA 2 | 50:50       | 30263Da          | ester   | 0.5-30kD-ester  |
| PLGA 3 | 60:40       | 42607Da          | ester   | 0.6-42kD-ester  |
| PLGA 4 | 60:40       | 33792Da          | acid    | 0.6-34kD-acid   |
| PLGA 5 | 85:15       | 61943Da          | ester   | 0.85-62kD-ester |

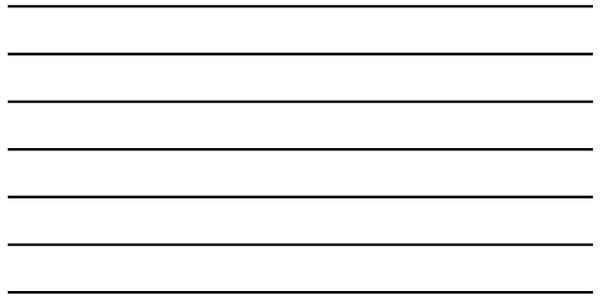
#### Printing conditions

| Composition     | 0.2 mm ID        |                |              | 0.4 mm ID        |                |              |
|-----------------|------------------|----------------|--------------|------------------|----------------|--------------|
|                 | Temperature (°C) | Pressure (bar) | Speed (mm/s) | Temperature (°C) | Pressure (bar) | Speed (mm/s) |
| 0.5-10kD-ester  | 110              | 9              | 1.5          | 95               | 7.5            | 1.5          |
| 0.5-30kD-ester  | 135              | 9              | 0.7          | 125              | 8              | 1.5          |
| 0.6-42kD-ester  | 165              | 9              | 0.7          | 140              | 9              | 0.5          |
| 0.6-34kD-acid   | 150              | 9              | 1.5          | 130              | 8              | 1            |
| 0.85-62kD-ester | 170              | 9              | 0.5          | 145              | 8              | 0.5          |

↑ @ 100 µm print resolution  
↓ @ 10 µm print resolution

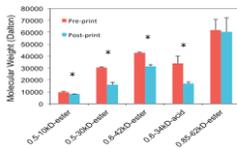
Slides courtesy Dr. Guo, TEBL

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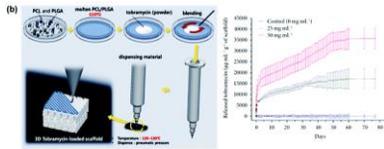
### Considerations: Bioink

- Material properties might alter due to a variety of reasons





## Homogenous immobilization



Gajjar, et al. Adv. Drug Delivery Rev. 2003.

Adapted from Biofabrication Workshop, Rice University




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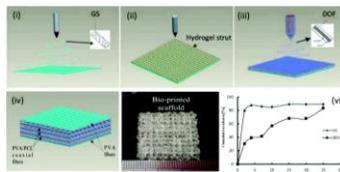
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## Heterogenous immobilization

- For temporal release of gentamycin sulfate and deferaxamine
- Blend electrospinning of polyvinyl alcohol 124-gentamycin sulfate (PVA-GS) fibers
- 3D printing for gelatin-sodium alginate struts
- coaxial electrospinning of PVA-DFO/PCL fibers



Liu, et al. Polym. Eng. Sci. 2016.

Adapted from Biofabrication Workshop, Rice University




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## Takeaways

- Several factors to be considered when
  - choosing the appropriate bioink
  - Printing procedure and impact on cells
  - Incorporating biological cues
- Post-processing impacts functionality just as much as pre- and during-
- Alternatives available, but have to be tailored to specific applications




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# Printing strategies and examples

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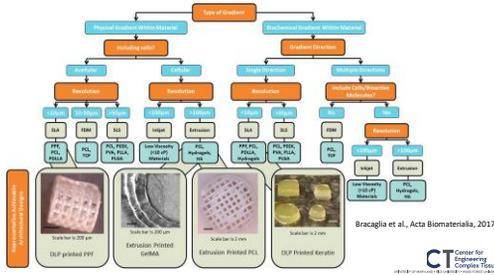
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## Choosing a 3D Printing Technique



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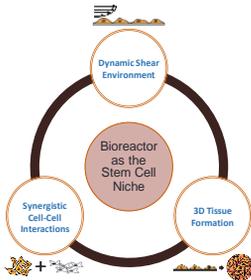
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## Bioreactors



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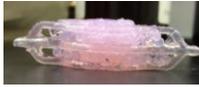
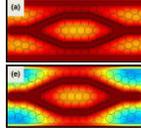
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### 3D Printed Vascular Network

3D Printed Template of a perfusable vascular network



Pore size and spacing affect nutrient exchange



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Ball et al., *Annals of Biomedical Engineering*, Vol. 44, No. 12, December 2016

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### Bioreactor Scale-up



BNN Nguyen, et al., *Tissue Engineering Part A*, 22: 263-271 (2016)

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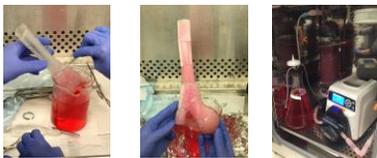
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### Bioreactor Scale-up



- 2.5 L osteogenic media flowing at 240 ml/min
- 250 mL of 2% alginate for cell encapsulation within 7200 alginate beads
- 1000mL of 2% alginate to fill empty space in culture chamber with 30,000 alginate beads
- Approximately  $800 \times 10^6$  hMSCs
- Sterilizing air filter on media flask to increase gas exchange

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BNN Nguyen, et al., *Tissue Engineering Part A*, 22: 263-271 (2016)

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### Bioreactor Scale-up



Volume of construct is **200 cm<sup>3</sup>**

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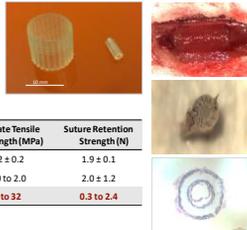
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### 3D Printed Vascular Grafts

- Grafts printed from poly(propylene fumarate) using a direct-light processing and crosslinked with UV light
- Mechanical properties similar to vessels used in autologous grafts



|                      | Modulus (MPa) | Ultimate Tensile Strength (MPa) | Suture Retention Strength (N) |
|----------------------|---------------|---------------------------------|-------------------------------|
| Human Saphenous Vein | 6.7 ± 1.3     | 2.2 ± 0.2                       | 1.9 ± 0.1                     |
| Human Femoral Artery | 9.0 to 12.0   | 1.0 to 2.0                      | 2.0 ± 1.2                     |
| 3D Printed Graft     | 11 to 176     | 1 to 32                         | 0.3 to 2.4                    |

Al Melchiorri, et al., *Advanced Healthcare Materials*, 5: 319-325 (2015).




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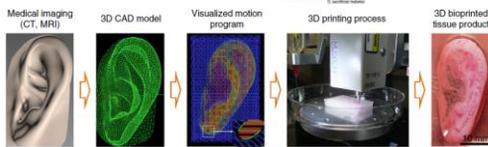
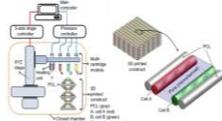
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### Custom-made, Multi-material platforms

#### Integrated Tissue-Organ Printer (iTOP)



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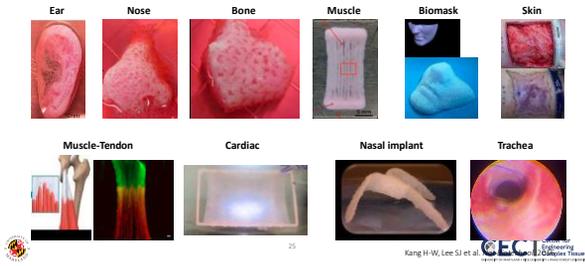
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### Custom-made, Multi-material platforms




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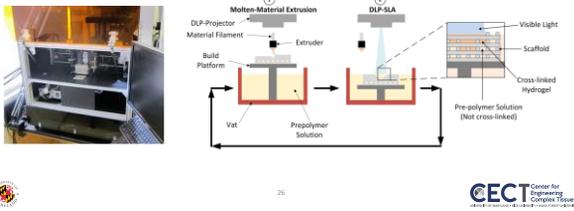
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### Custom-made, Multi-material platforms

#### Hybprinter

Y. Shanjani et al. 2015. *Biofabrication* 7 045008




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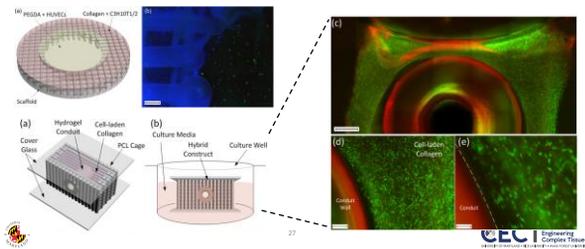
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### Custom-made, Multi-material platforms




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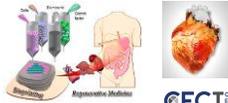
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## TR&D2: Live Cell Patterning

The ITOP can concurrently print synthetic biodegradable polymers and cell-laden hydrogels in a single tissue construct with clinically applicable size, shape, and structural integrity for clinical applications

- Generation of 3D freeform shaped constructs with precision
  - Multiple cell types, biomaterials, drugs
- High strength constructs
  - Gel and polymeric materials (~12)
- Printing resolution
  - Cell printing:  $\geq 50 \mu\text{m}$
  - Structural material printing:  $\geq 2 \mu\text{m}$




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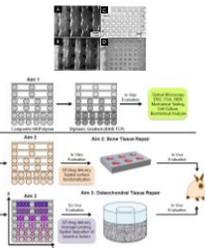
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## TR&D3: Complex Heterogeneous Scaffolds

- Develop a multi-material 3D printing system for the fabrication of complex bone and osteochondral scaffolds
  - Tunable material compositions
  - Patterned loading of growth factors
- Multi-material 3D printing system translatable to lower-cost 3DP systems
- Spatial deposition of transitional gradients (pore, ceramics, GFs) can mimic zonal organization
- Spatial manipulation of signaling properties to recapitulate tissue growth and regeneration in terms of composition and strength




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A NIBIB / NIH Biomedical Technology Resource Center  
Aiming to Grow the 3D Printing & Bioprinting Community

John Fisher (University of Maryland): 3D Printed Bioreactors for Dynamic Cell Culture  
 Antonios Mikos (Rice University): Bioprinting for Complex Scaffold Fabrication  
 Anthony Atala & James Yoo (Wake Forest University): Bioprinting for Cell-Laden Constructs

Center Collaborators: Jason Burdick (University of Pennsylvania), Elizabeth Cosgriff-Hernandez (Texas A&M University), Ali Khademhosseini (Brigham and Women's Hospital/Harvard), Helen Lu (Columbia University), David Mooney (Harvard University), Silvia Muro (University of Maryland), Anthony Ratcliffe (Synthasome), Molly Shochet (University of Toronto), Johnna Temenoff (Georgia Tech/Emory University), Rocky Tuan (University of Pittsburgh), Michael Yaszemski (Mayo Clinic), and Yunzhi Yang (Stanford University)




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## Acknowledgements

### Tissue Engineering & Biomaterials Laboratory

#### Lab Members

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Marco Santoro  
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