3t3 Cell Culture Protocol (ATCC CCL-98)

Characteristics –

Organism – mouse Tissue – embryo Cell type – fibroblast Growth Properties – adherent Biosafety Level - 1

Materials

- Performance FBS (Life Technologies)
- DMEM (ATCC, 30-2002)
- Pen/Strep (Life Technologies)
- Trypsin-EDTA (Life Technologies)

Media Formulation

- Complete Growth Media 1% PenStrep (v/v, 5 mL) + 10% (v/v, 50 mL) FBS + 89% (v/v, 445 mL) DMEM
- Cryoprotectant Medium Complete growth media + 5% (v/v) DMSO (1*10⁶ cells / mL)

Initiating Culture

- 1. Place a T-75 with complete growth medium into an incubator for at least 15 mins to allow the medium to reach its normal pH.
- 2. Thaw the vial by gentle agitation in a 37 C water bath (should take less than two mins)
 - a. note keep the O-ring and cap out of water
- 3. Transfer the vial contents to a centrifuge tube containing 9.0 mL complete culture medium, and spin at 600g for 5 mins
- 4. Resuspend cell pellet with the complete and dispense into the culture flasks from step 1.
- 5. Incubate the culture

Subculture

- 1. Remove and discard culture medium
- 2. Rinse the cells with PBS
- 3. Add 3mL (T-75) or 6mL (T-150) of Trypsin-EDTA solution and incubate at 37 C and observe after 5mins. Incubate for longer period of time until all the cells are lifted.
 - a. Do not agitate the cells by hitting or shaking the flask while waiting for cells to detach.
- 4. Add 6-8 mL of complete growth medium
- 5. Spin the cells at 600g for 5 mins and aspirate by gently pipetting
- 6. Resuspend cells in appropriate volumes of media
- 7. Add appropriate aliquots of the cell suspension to new culture flasks
 - a. A subcultivation ratio of 1:3 is recommended or 5 x 10³ cells/cm²
- 8. Incubate cultures at 37 C
- 9. Change the media twice a week.

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